



Asante Rogue Regional Medical Center (ARRMC)
Asante Three Rivers Medical Center (ATRCM)
Asante Ashland Community Hospital (AACH)

ANATOMIC PATHOLOGY

Anatomic pathology technical services are offered through Asante Laboratories. We contract with Vista Pathology, P.C., for professional, interpretive services.

For any questions regarding Anatomic Pathology, please call:

- **ARRMC & AACH: 541-789-4191, Monday-Friday, 8:00-5:00**
- **ATRCM: 541-472-7191, Monday-Friday, 8:00-4:30**

PATHOLOGY BILLING

Asante Laboratories processes anatomic specimens and submits them to Vista Pathologist for interpretation. Outpatients will usually receive a single bill from Vista Pathology which includes the charges for processing and interpretation of biopsies. Patients should be aware that on occasion they may receive two bills for Anatomic Pathology services; one bill from the hospital for preparation or special handling of specimens and one from Vista Pathology for interpretation. Charges for biopsies vary depending upon the number of submitted specimens and upon complexity of dissection and interpretation.

SUPPLIES/FORMS

Formalin-filled specimen containers and requisition forms are available through the Laboratory Courier Services office.

All specimens must be identified with the patient's name, birth date, and ordering physician, specimen source. Include a pre-op or post-op diagnosis and a brief clinical history when submitting pathology specimens. Biopsies must be accompanied by a duplicate billing requisition form. For outreach specimens, please provide adequate billing information to include the patient's name, age, birth date, address and copy of front and back of patient's insurance card(s). Including this information will save a phone call to your office.

SPECIAL INSTRUCTIONS

AUTOPSIES

Consult Pathologist: 541-789-4191 (Monday-Friday). On-call evening and weekend pathologist may be contacted with autopsy questions by calling:

- **ARRMC & AACH at 541-789-4164**
- **ATRCM at 541-472-7170**

HISTOLOGY SPECIMENS

Routine surgical specimens

Routine surgical specimens should be placed in 10% zinc formalin and submitted to pathology accompanied by a properly filled out requisition form. The formalin to specimen ratio should be 10:1. If not feasible (eg, large specimens) at least 4:1. Note: Fixation of tissue in formalin or other fixatives precludes certain special studies such as frozen sections and bacteriologic cultures. The time between removal of the specimen from the patient and immersion in formalin (cold ischemia time) must be available to the lab.

Rush Specimens

The department will prioritize specimens which are identified as rush. To order: write "RUSH" in **RED** on the tear-off portion of the form. Please note the reason for the rush status (medical – specify, nonmedical).

Specimens Requiring Special Handling

Specimens:

- Frozen Sections
- Bone Marrows
- Biopsy Cultures
- Muscle Biopsies
- Immunofluorescence
- Renal Biopsies
- Nerve Biopsies

CYTOLOGY SPECIMENS

The cytology department at Asante Rogue Regional Medical Center processes and evaluates both gynecologic (Pap test) and non-gynecological cytology specimens.

Cytology technical services are offered through Asante Laboratories. We contract with Vista Pathology, P.C. for professional, interpretive services.

For more information, please call the Cytology Department at 541-789-4195.

NOTE: Microslides made from the patient must be labeled with patient's name and source. All sample containers such as cardboard mailers, urine containers and liquid-based Pap sample vials etc. must be labeled with the PATIENT'S NAME AND DATE OF BIRTH. The sample must be accompanied by a matching requisition.

NON-GYNECOLOGICAL SPECIMENS (Non-pap Tests - e.g. Urine, Bronchial Washings, Body Cavity Effusions, Thyroid, LN Aspirations, etc.)

Cells disintegrate from enzymatic activity and, therefore, specimens should be sent to the Cytology Lab immediately after collection, or refrigerated until they can be sent to the Lab. If refrigeration is not possible, Saccomanno fixative is available from our Laboratory Outreach department.

DO NOT add formalin to Cytology specimens.

The usual turnaround time for nongynecological specimens is 24 hours after receipt, Monday through Friday. Billing for outpatient services and the professional component of hospital specimens is done through Vista Pathology, P.C.

NOTE: DO NOT add any fixatives if Microbiology testing is requested in *addition* to the Cytology test. Fixatives will kill any microorganisms present.

It is also very important not to reduce the volume of the Cytology specimen by pouring out any of the specimen to make room for the addition of fixatives. The preferred procedure would be to refrigerate the entire specimen until it can be sent to the lab. If this is not possible, then pour approximately one-half of the specimen into a second sterile container labeled with the patient's name, date of birth and specimen source, and then add fixative. Approximately 25% of the fluid in the container being fixative is sufficient. Send both containers to the lab.

Most specimens need only be handled as described above. The following specimens require special handling by the clinician.

Breast Aspirates

Prepare 2-4 smears like blood film smears however, **FIX IMMEDIATELY** with either 95% reagent alcohol or spray with hairspray as for a conventional PAP smear. An alternative would be to place the entire specimen in a sterile container (labeled with patient's name, date, time, and source); making sure specimen is completely covered with fluid. If not, add normal saline or Saccomanno fixative to prevent the cells from drying out. If only a small amount of specimen is obtained, aspirate a few mls of saline into the syringe, remove needle and cap syringe. Label syringe and send to lab with requisition slip as soon as possible.

Breast Discharge

Express fluid, wipe surface of nipple with soft gauze and express fluid again. Exfoliated cells are usually not obtained in the first fluid expressed from the breast Prepare 4-6 smears by placing glass slide against surface of nipple and draw across nipple to form a smear. **FIX IMMEDIATELY** in 95% Reagent Alcohol or spray with hair spray.

Bronchial Brushings

Clip end of the brush off into a properly labeled container with Saccomanno fixative or saline. Saccomanno Fixative is available from the Cytology department.

Cerebrospinal Fluid (CSF)

Cells tend to degenerate quickly in CSF, so it is especially important to keep these specimens refrigerated until they can be delivered to the lab.

Fine Needle Aspirations of Palpable Lesions (e.g., Thyroid, Salivary Glands, Lymph Nodes, etc.)

To arrange to have a Pathologist perform the fine needle aspiration of a palpable lesion, call RRMC Infusion Services Department 541-789-5006 or the Pathology Department at 541-789-4191 to schedule procedure.

Fine Needle Aspirations of Superficial Lesions Performed in the Office

Prepare up to ten smears, air dry approximately one-third and alcohol-fix the others by **IMMEDIATELY** placing smear in a screw cap container of reagent grade alcohol. Let the air-dried smears completely dry and send to lab in cardboard slide mailer or plastic slide transport container. Please label the air-dried smears with "A/D" in addition to patient's name and source.

Any specimen remaining in the needle and syringe should be rinsed in saline or Saccomanno fixative and submitted to the lab.

Urine

Due to degeneration of cells which occurs as the urine is in the bladder during the night, do not use the first early morning specimen. Hydrate the patient with 1 glass of water every 15 minutes for 1 hour. Send the entire first urine specimen following hydration to the Cytology Lab as soon as possible. Keep specimen refrigerated until Courier pickup to avoid cellular degeneration. Please specify on requisition slip how the specimen was collected (voided, clean catch midstream, ileal loop or catheterized). Include pertinent history such as the patient having a neobladder.

Sputum Specimen collection flyers are available from Laboratory Outreach department.

The best method of sputum collection **OTHER THAN BY INDUCTION**, is the early morning sputum. The Cytology Lab requests three early morning specimens on three separate days taken during the first hour after awakening. A pre-labeled container should be present at the bedside and the patient should have been instructed in the three-breath technique. This routine consists of three very deep breaths from the diaphragm which are held for five seconds each. The first two times the breath is exhaled with considerable pressure into a handkerchief or facial tissue which is held against the mouth. An explosive cough should accompany the third breath. The material produced at this time should be deposited in the sputum container. **DO NOT** put facial tissue or any type of fibrous material in with the specimen.

If the patient cannot bring the specimen to the Cytology Department within an hour of collection, the specimen should be refrigerated and brought to the Lab on ice or placed in 10-20 cc Saccomanno Fixative. A specimen which has been allowed to sit more than one hour unrefrigerated or unfixed may be unsatisfactory due to cellular degeneration. The best method of sputum collection is that which is induced by the Respiratory Department utilizing ultrasonic nebulization or aerosolized hypertonic saline. Arrangements for this type of collection may be made through the Respiratory Therapy Department.

NOTE: Patient's slide must be labeled with patient's name and specimen source. All sample containers such as cardboard mailers and liquid-based sample vials must be labeled with the patients' name AND DATE OF BIRTH. The sample must be accompanied by a matching requisition.

PAP TESTS

The Cytology department processes and evaluates both conventional PAP smears and liquid-based (SurePath) PAP specimens. Ancillary HPV testing and CT/NG testing can also be conducted from the liquidbased Pap specimens. Special instructions and brochures for the SurePath liquid-based Pap test can be obtained from our Laboratory Outreach department.

For conventional PAP specimens, place both the endocervical (non-hysterectomy patients) and ectocervical material on one slide and **IMMEDIATELY** spray fix with an alcohol-based fixative (we recommend hairspray). Allow smears to dry before packaging and sending to Lab.